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ISOLATION AND CHARACTERIZATION OF N-ALKYL-N-(BYDROXYMETHYL)NITROSAMINES FROM N-ALKYL-N-(HYDROPEROXYMETHYL) NITROSAMINES BY DEOXYGENATION

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Abstract: **N-Alkyl-N-(hydroxymethyl~nitrosmines, postulated intemediates in** the metabolic activation of carcinogenic nitrosamines, were prepared by deoxygenation of the corresponding hydroperoxymethyl nitrosamines and characterized.

N,N-Dialkylnitrosamines (1) are carcinogens and mutagens which require meta bolic activation. A postulated pathway of the activation ia **through a-hydroxy**lation." The intermediate N-alkyl-N-(l-hydroxyalkyl)nitrosamines (<u>2</u>) decompose spontaneously by heterolysis releasing aldehydes to provide reactive electrophilic species capable of alkylating nucleophiles. They are considered to be un **stable, and none of them has 80 far been isolated. Acylated derivatives of 2,**

/R Enzymatic OR-N $\begin{array}{ccccccc}\n\text{Enzymatic} & & \text{OR}-\mathbb{N} & & -\text{R}^t\text{CHO} & & & +\text{X}^-\n\end{array}$ **'CR2R' Hydroxylation CHR'** - N₂ (1)

N-alkyl-N-{l-acetoxyalkyl)nitroeamines (21, have been receiving considerable attention,² however, preparation of 2 from 3 was not successful.³ On the other hand, N-alkyl-N-(1-hydroperoxyalkyl)nitrosamines (4), prepared either by nucleophilic substitution of 3 with hydrogen peroxide in acetic acid⁴ or by oxygenatio **of a-lithiated 1. with oxygen, 5 may serva as a potential preoursor for the** pmparation of 2. This communication describes isolation, characterization and chemical properties of N-butyl- and N-methyl-N-(hydroxymethyl)nitrosamines, 2a and 2b, prepared from the corresponding hydroperoxymethyl nitrosamines by deoxy**genation.6**

In a typical preparative experiment, triphenyl phosphine (0.52 mmol) in CRC1₃ was added slowly to a CHC1₃ solution of $4a$ (0.30 mmol) while cooling with **ice under a stream of nitrogen. The sclution was concentrated m#ier reduced pressure and** the **concentrate was chromatograpbed** *on* **a columu of Sephadex LX-20** eluting with a mixture of CHT_{3} and methyl acetate $(9 : l)$ in an atmosphere of nitrogen. The fraction containing 2a was concentrated under reduced pressure an an appropriate aprotic solvent such as CH₃CN and DMSO was added to the concen**trated solution. The resulting solution** wasa **again concentrated to make** a solution in the desired solvent. Similarly, a solution of 2b was prepared from 4b **(0.20 mmol1 by treatamutwith** triphenyl **phosphine (0.34 mmel).**

2a and 2b were acetylated with acetic anhydride and pyridine at room temperature for 30 min to give 3a and 3b, respectively, whose identification was made **by comparison of the NRR and IR spectra with those of the authentic samples. 7** The yields for 3a and 3b from 4a and 4b were 92% and 90%, respectively, indica**ting that the deoxygenation reaction proceeded almost quantitatively.**

Sodium bisulfite can also be used for the deoxygenation. Aqueous bisulfite solution (1.34 M, 4.0 ml) was added to 4a (0.48 mmol), and after stirring the solution at 0° C for 1 min, the product was extracted with CHC1₃. It was purified by column chromatography as described above. Acetylation of the product 2a gave 3a in 81% yield calculated from $4a$. In a modified manner, 2b was prepared by passing a solution of 4b (0.38 mmol) in CHCl₃ through a column packed with a mixture of sodium bisulfite (7 g) and celite 545 (5 g) using CHCl₃ saturated with water as eluting solvent. After purification and acetylation, 3b was obtained in 48% yield from 4b,

a) (E):(2) ratio was determined by NMR integration of peak areas. b) ε -values were approxima<u>t</u>ely estimated on the basis of the concentration of acetate obtained by acetylation. c) disappeared on addition of D₂O. d) doublets changed to singlets by the addition of \bar{D}_2 0. e) disappeared on addition of CD₃OD. **f) doublets changed to jsinglets by the addition of CD30D. g) indistinguishable** under the condition examined.

The NMR and UV spectral data for 2a and 2b are shown in the table. The hydroxymethyl **structure waa supported by** *comparison* **of their** chemical **shifts observed in the NRR** spectra with those **of the** corresponding acetoxymethyl, methoxymethyl⁷ and hydroperoxymethyl compounds.⁴ The structure was further confirmed on the basis **of the effect of deuterium exchanges.**

The rate of decomposition in sodium phosphate solution with a constant ionic strength of 0.2 in the pH range 1 -8 was calculated from the time dependency of the decrease of logarithmic values of the UV absorption at 228 nm due to N-NO function, using the least squares method. \bullet for $2a$, and o for $2b$.

The hydroxymethyl compounds were unstable in aqueous solutions. The rate constant of the decomposition is plotted as a function of pH of the aqueous phosphate solution, as shown in the figure. No significant difference was observed between $2a$ and $2b$. They were unstable in neutral as well as in alkaline solution, and a linear increase in the rate constant was observed at pH above 6 . While, they were rather etable in acidic media, having a half-life of 5.4 min and 5.9 min for $2a$ and $2b$, respectively, at pH 2 to 4. In ethanol, they decomposed with a similar half-life of 5.8 min and 5.7 min for 2a and 2b, respectively. The hydroxymethyl compounds were stable enough to make the isolation possible in carefully dried aprotic solvents as described above.

According to the postulated metabolic pathway, the α -hydroxy intermediate decomposes to yield an aldehyde and an alkylating species, which can be trapped by water in aqueous Solution. Formaldehyde, and isomeric butanols or methanol were identified from 2a or 2b, respectively. Formaldehyde was isolated as its 2,4-dinitrophenylhydrazone and identified by comparison of the NMR and IR spectra with those of the authentic sample. The alcohols were detected by gas chromatography and identified by comparing their retention time with those of the authentic samples on two different columns. ⁸ Thus a solution of the hydroxymethyl compound in 0.5 ml of CH₃CN or DMSO was treated with 10 ml of water for 1 h at room temperature. **A** portion **of** the solution was then treated with 2,4-dinitrophenylhydrazine reagent, and another portion was diluted with water and analyzed directly by gas chromatography. 2a (0.13 mmol) yielded formaldehyde, 1-butanol and 2-butanol in 81%, 44% and 20% yields, respectively. 2b (0.26 mmol) gave formaldehyde and methanol in 73% and 93% yields, respectively.

3695

In the presence of thiophenol, a nucleophile other than water, its methylation or butylation was observed with 2b or 2a, respectively. 2b (0.66 mmol) was **treated at room temperature with potassiumthiophenolate (5 mmol) in 200 ml of** sodium **phosphate buffer (pH 7.4) with an ionic strength of 0.2 for 1 h. After purification of the product by column chromatography, methyl phony1** nulfide **was obtained in 26% yield and identified by NMR and IR spectroscopies. Under a simi**lar condition, 2a (0.75 mmol) gave butyl phenyl sulfide in 1.2% yield.

The result described here presents the first example of the **isolation and characterization of the postulated unstable intermediate involved in the metabolic activation of carcinogenic dialkylnitrosamines. The deoxygenation reaction with primary hydroperoxy derivativee of dialkylnitrosamines proceeded almost quantitatively. The decomposition pathway** of the key **intermediate was also elucidated, and the alkylating activity of the intermediate was clearlydemonstrated, thus** subetantiating the **postulated metabolic activation pathway of carcinogenic dialkylnitrosamines.**

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Pack 54, 60-80 mesh (3.1 m × 3 mm) at 90°C, retention time, 7.6 min (methano **pack 54, 60-80 mesh, (3.1 m x 3 nun) at 9O"C, retention time, 7.6 min (methanol),** 12.4 min (acetaldehyde, internal standard); qualitative analysis: 15% PEG **1000 on Shimalite F; 20-80 mssb (1.5 m x 3 mm) at BO*C, retention time, 2.8** min (methanol), 5.8 min (2-butanol), 12.6 min (1-butanol).

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